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ORIGINAL
RESEARCH

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BACKGROUND AND PURPOSE: DWI by using higher b-values provides tissue diffusivity with less T2 shine-through effect. VCD in the abscess cavity correlates with ADC values. The purpose of this study was to investigate which b-value–derived ADC correlates better with VCD.

MATERIALS AND METHODS: Thirty patients with brain abscess underwent conventional MR imaging and DWI with $b=1000$, 2000 , and 3000 s/mm² on a 3T MR imaging scanner. ADC values were quantified by placing regions of interest inside the abscess cavity in all sections where the lesion was apparent on coregistered ADC maps derived from different b-values. VCD was measured on pus aspirated.

RESULTS: An increase in b-value was associated with a decrease in ADC values in normal parenchyma as well as in the abscess cavity. The most significant negative correlation of VCD was observed with $b=3000$ s/mm² ($r = -0.98$, $P = .01$).

CONCLUSIONS: VCD in the abscess cavity can be best assessed at $b=3000$ s/mm² secondary to the reduction in the T2 shine-through effect. DWI with $b=3000$ s/mm² is of promising value in the assessment of the therapeutic response of brain abscess.

ABBREVIATIONS: ADC = apparent diffusion coefficient; CNWM = contralateral normal white matter; DWI = diffusion-weighted imaging; FLAIR = fluid-attenuated inversion recovery; FWM = frontal white matter; GM = gray matter; spp = species; VCD = viable cell density; WM = white matter

Brain abscess is a focal suppurative process within the brain parenchyma.¹ The incidence of brain abscess is highly variable. Brain abscesses comprise 1%–2% and up to 8% of all the patients with intracranial space-occupying lesions seen in the developed and developing countries, respectively.² Despite significant advances in diagnosis and treatment, brain abscess remains a potentially fatal central nervous system infection.³ The abscess cavity contains inflammatory cells with intact cell membranes and nuclei, necrotic tissue debris, proteinaceous exudates, bacterial metabolites, and cytosolic amino acids, with or without the brain abscess's causative organism.⁴

CT and conventional MR imaging are the main noninvasive modalities used to diagnose brain abscess. However, the imaging features of brain abscesses are nonspecific and may simulate those of cystic rim-enhancing mass lesions of varying etiologies.⁵ DWI is widely used in the detection and character-

ization of a large number of brain pathologies, such as stroke, abscess, encephalitis, and tumors involving the central nervous system.^{6,7} DWI has been shown to be useful in distinguishing brain abscess from necrotic or cystic brain tumors.^{8,9}

The restriction of diffusion in brain abscess is mainly influenced by the attenuation of viable inflammatory cells present in the pus, irrespective of the etiology of the abscess.¹⁰ Treated brain abscesses have higher ADCs compared with untreated abscesses, and return to low ADCs suggests reactivation of the disease.¹¹ Recently a few studies have shown the utility of high b-values in the diagnosis of various disease conditions.^{12–19} Theoretically, higher b-value DWI provides better contrast with its reflection of more tissue diffusivity and less T2 shine-through effect.^{13,14} However, at 1.5T or lower field strengths, higher b-values are not usually used in clinical practice due to inferior signal intensity-to-noise ratios.^{15–17}

It has been shown in vitro that the magnitude of diffusion in intracellular water molecules is an order smaller than that in the extracellular space.^{20,21} The water diffusion in biologic tissues can be described in terms of 2 main components: water molecules of low mobility with slow diffusion (either bound to macromolecules or confined within the cell membrane) and of high mobility with high diffusion (mostly extracellular).^{22,23} The acquisition of DWI up to $b=4000$ s/mm² amplifies the sensitivity of DWI to assess water diffusion properties and enables the separation of signals arising from the intracellular and extracellular water molecules.^{20,21} Hence, DWI should be sensitive to several physiologic and morphologic characteristics of the tissues, which are associated with the diffusion of the low and high mobility of water molecules. Two water compartments separated with different diffusion coefficients (ie, extracellular water for the compartment with high

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ADC) have been explained on the basis of the biexponential fit model for water signal-intensity decay with different b-values.²⁰⁻²² This biexponential diffusion property of water has been used in functional task-activation assessments.²⁴

The purpose of this study was to investigate which b-value—derived ADC correlates better with VCD. We computed a biexponential fit to understand which component is influenced by the high b-value in the abscess cavity.

Materials and Methods

Study Design

Thirty-five patients with suspected intracranial abscess (24 males and 11 females; mean age, 25 years; ranging from 9 to 47 years of age) admitted to the Department of Neurosurgery, Chhatrapati Sahuji Maharaj Medical University, Lucknow, India, from August 2009 to December 2010 and 10 healthy controls (ranging from 23 to 45 years of age) were enrolled in the study prospectively. These subjects underwent conventional MR imaging and DWI in the Department of Radiodiagnosis, Sanjay Gandhi Post Graduate Institute of Medical Sciences, Lucknow, India, on a 3T MR imaging scanner (Signa Hdx; GE Healthcare, Milwaukee, Wisconsin) with a 12-channel head coil. Institutional ethics approval and informed consent from all patients or their nearest kin were obtained before the investigation. Diagnosis of brain abscess was suggested on the basis of diffusion imaging in 30 patients and formed the basis of inclusion for the study protocol design. The diagnosis was confirmed in all these 30 patients after surgery.

MR Imaging Protocol

The conventional MR imaging protocol included T2-weighted fast spin-echo with TE/TR/NEX = 88/4400 ms/1.2; number of sections = 42; section thickness = 3 mm; FOV = 240 mm; T2 FLAIR with TE/TR/NEX = 140/9000 ms/2; number of sections = 42; section thickness = 3 mm; FOV = 240 mm; and spin-echo T1 FLAIR with TE/TR/NEX/flip angle = 14/1300 ms/1/65°; number of sections = 42; section thickness = 3 mm; FOV = 240 mm with image matrix = 256 × 256 and no interslice gap.

All patients underwent DWI at $b=1000, 2000$, and 3000 s/mm^2 . DWI parameters were the following: TE = 93.2 ms; TR = 7075 ms; NEX = 2.0; number of sections = 42; section thickness = 3 mm; FOV = 240 mm; image matrix = 128×128 with no interslice gap. Postcontrast T1-weighted images were acquired after intravenous injection of gadadimide dimeglamine (Omniscan, Oslo, Norway) at a dose of 0.1 mmol/kg of body weight.

Culture of Abscess Aspirate

All patients underwent surgery within 24 hours of imaging. The pus was collected in a sterilized vial and snap-frozen in liquid nitrogen immediately after surgery/aspiration. At the same time, 1–2 mL of aspirated pus was inoculated into BACTEC Plus Aerobic/Anaerobic media (Becton Dickinson, Sparks, Maryland) to isolate the aerobic, facultative anaerobic, and anaerobic bacteria for the confirmation of the diagnosis of etiologic agents. The inoculated media were incubated at 37°C, and growth was monitored in BACTEC 9050 (Becton Dickinson) for 5 days. Media with positive growth were subcultured on appropriate solid media and incubated aerobically and anaerobically at 37°C. Anaerobic incubation was carried out in jars filled with a gas mixture of nitrogen gas (80%–90%), carbon dioxide (5%–10%), and deuterium (5%–10%), through an Anoxomat system (Mart Mi-

crobiology, Drachten, the Netherlands). All of the isolates were identified by standard biochemical tests as described previously.²⁵ Cultures and smear examinations were performed for *Mycobacterium tuberculosis* as well as for fungi in all cases.²⁶

Viable Cell Count

In this study, the viability of the cells was calculated by a hemocytometer (Burker-Turk; Erma, Tokyo, Japan) after diluting pus 100- to 1000-fold in white blood cell diluting solution and expressed as cells/milliliter as described previously.¹⁰ Inflammatory cells (leukocytes) with intact cell membranes and nuclei, as identified by 0.5% trypan blue staining, were referred to as viable cells. The method for the calculation of cell attenuation has been described elsewhere.^{10,27}

Data Analysis

A Java-based Image J plug-in (<http://rsb.info.nih.gov/ij>) was developed to generate the ADC maps from DWI datasets with different b-values.

Biexponential Fit and Its Quantification

The diffusion-weighted signal intensity in case of biexponential decay is given by

$$1) \quad S(b) = S(0)(f_{fast}\exp[-b \cdot D_{fast}] + f_{slow}\exp[-b \cdot D_{slow}]),$$

where $S(b)$ is the signal intensity in the presence of a diffusion gradient, $S(0)$ is the signal intensity in the absence of a diffusion gradient, D_{fast} and D_{slow} are the fast and slow ADCs, and f_{fast} and f_{slow} represent the respective fractional contributions to the signal intensity. With $a_{fast} = S(0) - a_{slow} = S(0)f_{fast}$, equation 1 can be written as

$$2) \quad S(b) = a_{fast}(\exp[-b \cdot D_{fast}] - \exp[-b \cdot D_{slow}]) + S(0)\exp(-b \cdot D_{slow}).$$

Coregistration was done between datasets with different b-values before fitting of equation 2 to obtain the ADCs of fast and slow diffusing components. Also the conventional ADC maps were calculated by using the monoexponential diffusion equation, $S(b) = S(0)\exp(-b \cdot D)$ from the above coregistered datasets with different b-values from patients as well as controls.²⁸ The mean ADCs, ADC_{fast} , and ADC_{slow} components were calculated by placing elliptic or circular regions of interest ($40\text{--}60 \text{ mm}^2$) on any 1 of the above maps (and applying the same region of interest on other maps) inside the abscess cavity in all sections where the lesion was present. Similarly mean ADC values were also quantified from the CNWM and GM regions of the each patient with brain abscess in each section as they were from the abscess cavity.

Statistical Analysis

Bivariate analysis of the Pearson correlation was performed to study the relationship between mean ADC values, fast and slow ADC values, and VCD quantified from the abscess cavity. An independent Student *t* test was also performed to see any changes in ADC values quantified from the abscess cavity, CNWM, and GM regions of patients with brain abscess as well as FWM and GM regions of healthy controls at different b-values. All statistical analyses were performed by using the Statistical Package for the Social Sciences, Version 16.0 (SPSS, Chicago, Illinois).

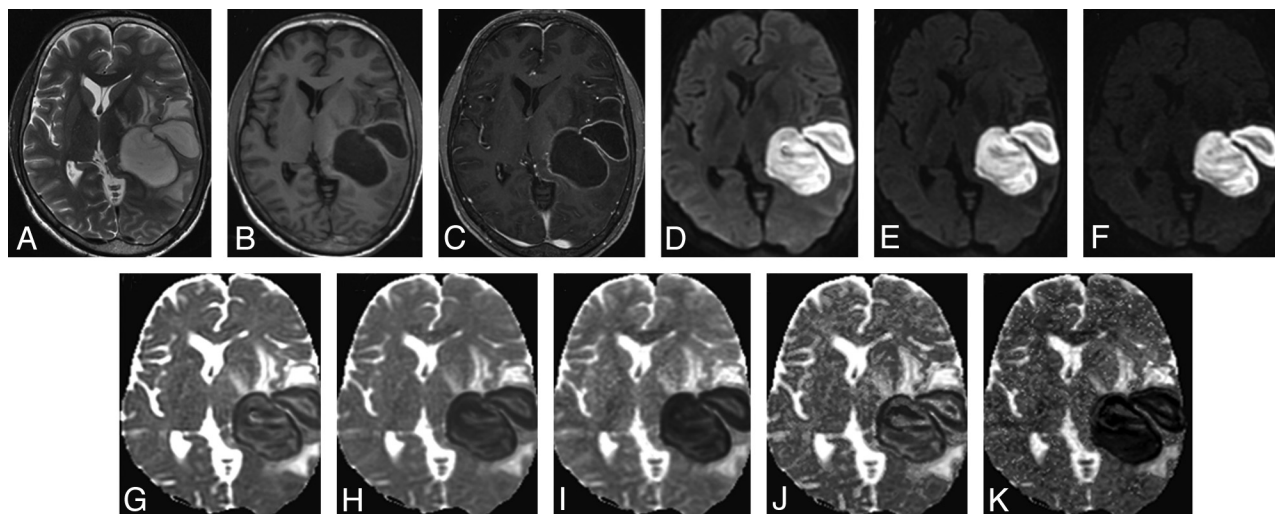


Fig 1. A 16-year-old male patient with brain abscess. *A*, T2-weighted image shows bilobed centrally hyperintense lesions with a peripheral hypointense rim and surrounding edema in the left temporo-parietal region causing mass effect. *B* and *C*, The lesion shows central hypointensity with a peripheral isointense rim on the T1-weighted image (*B*) and rim enhancement on the postcontrast T1-weighted image (*C*). *D–F*, DWI at b-values of 1000 (*D*), 2000 (*E*), and 3000 s/mm² (*F*), respectively, show intense hyperintensity inside the lesions with maximal suppression of free water at 3000 s/mm² (*F*). *G–I*, ADC maps generated from DWI at different b-values (1000, 2000, and 3000 s/mm²) show a decrease in signal intensity inside the lesions with increasing b-values, suggestive of maximal suppression of extracellular water. *J* and *K*, The fast (*J*) and slow (*K*) component ADC maps derived from the biexponential fit in the abscess cavity have values of $(1.11 \pm 0.06) \times 10^{-3}$ s/mm² and $(0.15 \pm 0.23) \times 10^{-3}$ s/mm², respectively. Pus culture showed *S aureus*.

Table 1: ADC values quantified from the abscess cavity, CNWM, and GM of patients with brain abscess and the WM and GM of healthy controls at different b-values

	ADC ^a			
Regions	A (<i>b</i> =1000)	B (<i>b</i> =2000)	C (<i>b</i> =3000)	<i>P</i> Value
Patients (<i>n</i> = 30)				
Abscess cavity	0.59 ± 0.20	0.48 ± 0.13	0.39 ± 0.11	A vs B = .05; A vs C = .01, B vs C = .05
CNWM	0.64 ± 0.04	0.63 ± 0.11	0.62 ± 0.14	A vs B = .86, A vs C = .80, B vs C = .93
GM	0.74 ± 0.04	0.71 ± 0.12	0.71 ± 0.02	A vs B = .37, A vs C = .01, B vs C = .97
Healthy controls (<i>n</i> = 10)				
WM	0.82 ± 0.02	0.68 ± 0.02	0.58 ± 0.02	A vs B = .01, A vs C = .01, B vs C = .01
GM	0.78 ± 0.03	0.71 ± 0.01	0.64 ± 0.02	A vs B = .01, A vs C = .01, B vs C = .01

^a(Mean ± SD) × 10⁻³ s/mm².

Results

Clinical and Laboratory Examination

The predisposing factors were recognized in 24 cases: otitis media (n = 15), congenital heart disease (n = 3), postoperative infection (n = 1), steroid therapy (n = 2), head injury (n = 1), and pulmonary tuberculosis (n = 2). In the remaining 6 patients, the source of infection could not be ascertained. Of a total of 30 brain abscesses, 18 were found to be pyogenic, 1 was fungal, and 3 were confirmed as tubercular on a culture of aspirated pus. The micro-organisms isolated on culture and confirmed by standard biochemical test were aerobic *Streptococci* spp (n = 10), *Staphylococcus aureus* (n = 1), *Bacteroides* spp (n = 2), *Nocardia* spp (n = 1), *Histoplasma capsulatum* (n = 1), mixed microbes (aerobic and anaerobic) (n = 4), and tubercular (n = 3). Eight cases were sterile and were considered pyogenic due to the presence of acute inflammatory cells and the absence of *Mycobacterium* spp and fungal growth on microscopy and culture. In the abscess cavity, the VCD varied from 6000 to 11,000 cells/mm³.

Imaging Findings

Brain abscesses were located in the frontal lobe (n = 3), parietal lobe (n = 4), temporal lobe (n = 10), parietotemporal

lobe (n = 4), and cerebellum (n = 9). The brain abscess appeared hyperintense on T2-weighted images with a peripheral hypointense rim (Fig 1A) and iso- to hypointense on T1-weighted images (Fig 1B). Postcontrast study showed rim enhancement (Fig 1C). A few brain abscesses showed homogeneous hyperintensity on DWI (Fig 1D–F), with corresponding homogeneous hypointensity on ADC images (Fig 1G–I). However, most brain abscesses showed heterogeneity on DWI and corresponding ADC images.

Quantification of ADC

The mean ADC values of the abscess cavity, CNWM, and GM regions quantified by placing regions of interest on ADC maps obtained from different b-values are summarized in Table 1. On independent Student *t* tests, a significant decrease in mean ADC values was observed in the abscess cavity at different b-values. In CNWM, there was no significant difference among mean ADC values obtained from different b-values. In GM regions, there was a significant difference only between mean ADC values of b=1000 and b=3000 s/mm² (Table 1). In healthy controls, we observed significant differences in mean ADC values quantified from FWM and GM regions at different b-values (Table 1).

Table 2: Mean values of fast and slow components of ADC quantified from the abscess cavity, CNWM, and GM at different b-values of patients with brain abscess

Group	ADC ^a			P Value
	Abscess Cavity (A)	CNWM (B)	GM (C)	
Fast	1.11 ± 0.06	1.03 ± 0.13	1.12 ± 0.12	A vs B = .62, A vs C = .95, B vs C = .06
Slow	0.15 ± 0.23	0.26 ± 0.10	0.30 ± 0.02	A vs B = .09, A vs C = .01, B vs C = .1

^a(Mean ± SD) × 10⁻³ s/mm².

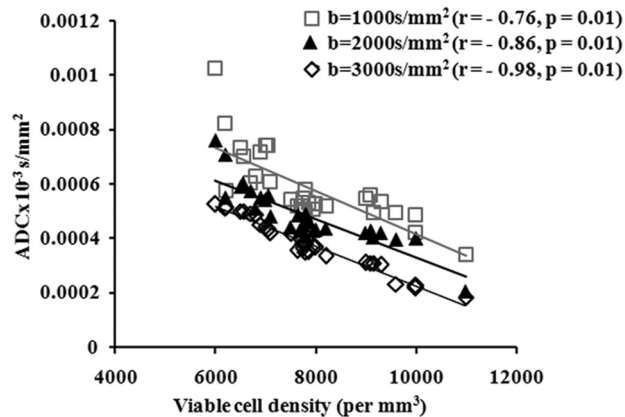


Fig 2. Scatterplot shows correlation between VCD with mean ADC values measured from different b-values ($b=1000$, 2000 , and 3000 s/mm²) within the abscess cavity of patients with brain abscess.

On independent Student *t* tests, no significant difference was observed between ADC_{fast} of brain abscess lesions, CNWM, and GM regions of patients with brain abscess. ADC_{slow} of the abscess cavity was found to be significantly lower than the ADC_{slow} of the GM of patients with brain abscess (Table 2).

On bivariate analysis by using the Pearson correlation, a significant negative correlation was observed between the VCD and mean ADC values from different b-values ($b=1000$ s/mm², $r = -0.76$, $P = .01$; $b=2000$ s/mm², $r = -0.86$, $P = .01$; and $b=3000$ s/mm², $r = -0.98$, $P = .01$) (Fig 2). No significant correlation was observed between VCD and fast and slow ADC components computed through biexponential fit (ADC_{fast} [$r = -0.17$, $P = .56$] and ADC_{slow} [$r = -0.15$, $P = .58$]).

The biexponential fit of combined and fast components revealed a decrease in signal intensity with increasing b-values in the WM and GM regions and abscess cavity (Fig 3A–C). However, there was a gradient in decrease in the signal intensity of the slow component moving from GM to WM regions with not much change in the abscess cavity as a function of b-values (Fig 3A–C).

Discussion

In this study, significantly low mean ADC values were observed in the abscess cavity at different b-values. With increasing b-values, ADC values decreased in the abscess cavity, GM, and CNWM regions of brain. The decrease in ADC values

between $b=1000$ and $b=3000$ s/mm² was significant in the GM of patients and controls, while it also showed only a significant decrease from FWM in healthy controls. Although there was a significant negative correlation of VCD with all 3 b-values, the correlation at $b=3000$ s/mm² was found to be close to 1. The quantification of fast and slow components of ADC by using a biexponential fit in the abscess cavity did not correlate with VCD.

It has been shown by in vivo and ex vivo experiments that the increase in cell attenuation in brain abscesses negatively correlates with mean ADC values.¹⁰ The mean ADC values measured from the abscess cavity showed trends similar to those exhibited by cellular tumors in humans²⁶ and animal tumor models.²⁷ It has been reported that b-value is inversely proportional to ADC.²⁹ DeLano et al¹⁴ have reported that when b-values increase from 1000 to 3000 s/mm², ADC values decrease from 30% to 35% for the same regions of interest. The authors related it to the biexponential decay in signal intensity. The signal-intensity decay observed in their study as a function of b factor is in close agreement with our results from a biexponential model in the abscess cavity of these patients (Fig 3A–C).

It has been reported that the ADC of viable tissue is a result of independent contributions from the intracellular and extracellular compartments.^{30,31} Therefore, at least 2 factors contribute to a decrease in ADC values with increasing cell attenuation. The first is that tissues with high cell attenuation have a large fraction of intracellular water compared with tissues with low cell attenuation. The intracellular water has presumably lower ADC than the extracellular water due to a higher content of diffusion barriers in the intracellular compartment.³⁰ Second, tissues with high cell attenuation have small extracellular compartments compared with tissues with low cell attenuation. The mobility and mean ADC decrease in tissues with high cellularity results in low ADC because of small extracellular water content. The influence of this factor is strengthened because the ADC is biased toward the diffusion coefficient of the compartment with the longest T2 (ie, the extracellular compartment).³¹

On the biexponential fit model, we observed a decrease in signal intensity of the fast component as we moved from low to high b-values. The strongest correlation of VCD with ADC values at $b=3000$ s/mm² can be attributed to the fact that on increasing diffusion weighting, there is a decrease in T2 shinnethrough effect, resulting in removal of fast components, which in turn is responsible for more precise measurements of ADC values of the intact inflammatory cells inside the abscess cavity. The absence of any change in signal intensity of the slow component inside the abscess cavity further suggests that mainly the fast component is probably responsible for the T2 shinnethrough effect, which is removed on increasing the diffusion-weighting.

On the basis of biexponential fit, the fast and slow diffusion components of water have been described in human brain and interpreted as from extracellular and intracellular compartments, respectively.^{32,33} Le Bihan²⁴ has suggested that fast and slow components of the water diffusion correspond to 2 differently structured water pools, rather than specific water compartments. He proposed a model where both the slow and fast components of water diffusion originate partly in the in-

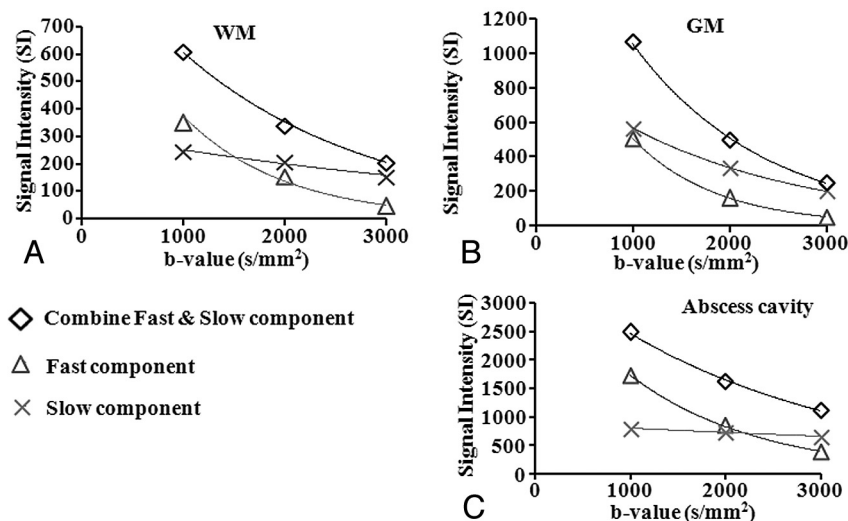


Fig 3. Graph shows biexponential fitting in the WM (A), GM (B), and abscess cavity (C) of patients with brain abscess. On increasing the b-value, there is a reduction in the signal intensity of combined ADC values and the fast ADC component in WM, GM, and the abscess cavity, but slow ADC component shows gradual reduction from GM to WM with not much change in the abscess cavity.

tracellular space and partly in the extracellular space.²⁴ Although we observed the best correlation with ADC at $b=3000$ s/mm², we did not observe any correlation of fast and slow components of the ADC with the actual cell count, suggesting that these 2 components may not represent the extra- and intracellular water compartments, respectively. Our results further confirm that these 2 components of water diffusion originate from the 2 different pools rather than from the 2 different compartments of water.

The viable inflammatory cells in the abscess cavity are the result of active inflammation, and a decrease in inflammatory cells is considered a marker for the response to therapy. This has been shown by serial ADC measures in which an increase in ADC indicates a response, while a decrease in its value suggests reactivation of the disease.¹¹ Similar results have been shown by using serial fractional anisotropy measurements in brain abscess, in which an increase in fractional anisotropy is associated with upregulation of inflammatory cytokines and increased inflammatory cells, while the reverse is true when there is a positive response.^{34,35} Usiskin et al³⁶ have also used serial high b-values (up to $b=3000$ s/mm²) to assess the response to highly active antiretroviral therapy in a case of progressive multifocal leukoencephalopathy. Our data suggest that ADC maps derived from $b=3000$ s/mm² may be ideal for quantification of intact inflammatory cells in the abscess cavity and may be able to predict therapeutic response much better than ones obtained from $b=1000$ s/mm².

Various authors have used b-values up to 5000 s/mm² to demonstrate the biexponential fit in human brain.^{22,23,29} In the current study, we used b-values up to 3000 s/mm², which may be suboptimal to demonstrate the biexponential fit and may be considered a limitation.

Conclusions

The viability of cells, a measure of disease activity, can be best quantified by ADC values derived by using $b=3000$ s/mm² DWI. This may be used in the future to assess the therapeutic response to antibiotics in patients with brain abscess. The fast

and slow components of diffusivity may not truly represent the extracellular and intracellular compartments and may simply represent the free and bound water present in the tissue.

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